- Crampin, S. Geological and industrial implications of extensive-dilatancy anisotropy. *Nature* 328, 491–496 (1987).
  - Sobolev, S. V., Babeyko, A. Y. & Garfunkel, Z. Desert Group. Thermo-mechanical model of the Dead Sea Transform. *Earth Planet. Sci. Lett.* (submitted).
  - Wolfe, C. J., Vernon, F. L. III & Al-Amri, A. Shear-wave splitting across western Saudi Arabia: The pattern of upper mantle anisotropy at a Proterozoic shield. *Geophys. Res. Lett.* 26, 779–782 (1999).
  - Reinecker, J., Heidbach O. & Müller, B. The 2003 release of the World Stress Map (http://www.worldstress-map.org/) (2003).
  - Bartov, Y., Avni, Y., Calvo, R. & Frieslander, U. The Zofar fault A major intra-rift feature in the Arava Rift Valley. Geol. Surv. Israel Curr. Res. 11, 27–32 (1998).
  - Savage, M. K. Seismic anisotropy and mantle deformation: What have we learned from shear wave splitting? *Rev. Geophys.* 37, 65–106 (1999).
  - McNamara, D. E., Owens, T. J., Silver, P. G. & Wu, F. T. Shear wave anisotropy beneath the Tibetan Plateau. J. Geophys. Res. 99, 13655–13665 (1994).
  - Hirn, A. et al. Seismic anisotropy as an indicator of mantle flow beneath the Himalayas and Tibet. Nature 375, 571–574 (1995).
  - Herquel, G., Tapponnier, P., Wittlinger, G., Mei, J. & Danian, S. Teleseismic shear wave splitting and lithospheric anisotropy beneath and across the Altyn Tagh fault. *Geophys. Res. Lett.* 26, 3225–3228 (1999).
  - Nicolas, A. & Christensen, N. I. in Composition, Structure and Dynamics of the Lithosphere-Asthenosphere System (eds Fuchs, K. & Froidevaux, C.) Geodyn. Ser. 16, 111–1123 (1987).

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# The oldest articulated chondrichthyan from the Early Devonian period

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Chondrichthyans (including living sharks, skates, rays and chimaeras) have a fossil record of scales and dermal denticles perhaps dating back to the Late Ordovician period, about 455 million years ago<sup>1,2</sup>. Their fossil tooth record extends to the earliest Devonian period, almost 418 million years ago<sup>3</sup>, whereas the oldest known articulated shark remains date from the Early Devonian period<sup>4</sup>, about 394 million years ago<sup>5</sup>. Here we report the discovery of an articulated shark that is almost 409 million years old<sup>5</sup> from the Early Devonian (early Emsian) period of New Brunswick, Canada. The specimen, identified as *Doliodus problematicus* (Woodward)<sup>6</sup>, sheds light on the earliest chondrichthyans and their interrelationships with basal jawed **cartilaginous fishes.** Isolated and articulated Early to Middle Devonian shark specimens are rare<sup>1,9</sup>. Until now, the oldest partial articulated shark, consisting of the braincase articulated with parts of the visceral skeleton, was *Pucapampella* from the Early Devonian of South Africa<sup>4</sup>. Significant Middle Devonian partially articulated specimens include *Pucapampella* from Bolivia<sup>10,11</sup>, *Antarctilamna prisca* from Antarctica and Australia<sup>7,12</sup>, and *Gladbachus adentatus* from Germany<sup>13</sup>.

Specimen NBMG (New Brunswick Museum, Geology) 10127/ 1a,b-4 consists of the anterior part of *D. problematicus*, forward of the mid-trunk region (Fig. 1). It is preserved dorsoventrally, oriented dorsal side up with exo- and endoskeletal elements preserved, including characteristic prismatic calcified cartilage, teeth, scales and large fin-spines. The specimen is cleaved in five parts, providing a series of transverse sections through the head and branchial region. The preserved length is 23 cm, suggesting a body length of perhaps 50–75 cm on the basis of shark comparative anatomy.

Prismatic calcified cartilage, considered to be a chondrichthyan synapomorphy<sup>1,14</sup>, compose the neurocranium and splanchnocranium. The articulated jaws confirm that D. problematicus possessed tooth families and provide early evidence in chondrichthyans of the relationship of teeth to the dental lamina<sup>1,15</sup>. Most teeth are partially buried; however, tooth families that are visible have teeth stacked in a row, with newer teeth sitting in a space representing the position of the dental lamina groove. Tooth bases abut a prominent darkbrown concave surface, interpreted as preserved basal connective tissue. The dentition shows weak dignathic and disjunct monognathic heterodonty, suggesting revision of earlier opinions about the evolution of shark teeth<sup>16</sup>. Functional upper and lower teeth, offset anteriorly, oppose one another with sharp lateral edges of principal cusps connecting in a scissors movement. The functional teeth show the asymmetry and range of variation previously recognized<sup>2,6,8</sup>, and verify the position and number of tooth types in the jaw. Teeth are not seen in the symphysial and parasymphysial portions of the lower jaw.

The right side of the lower jaw shows about 15 tooth families; the left side has only 11 tooth families preserved, with bases of at least three anterior rows present in the cartilage. Tooth families expose up to three teeth each. Near the posterior jaw articulation, flat basal pads might represent the most posterior teeth. Lower tooth families are seen in cross-section, showing the apparently highly vascularized lower edge and new tooth germs. The last three to four posterior tooth families do not show dental membranes and thus are more like modified dermal scales. In a few teeth, two large divergent lateral outer cusps with two to four intermediate small cusps can be seen in cross-section. These and a thin section of a D. problematicus tooth from the National Museums of Scotland (RSM1897.51.46) show that the cusps are formed of orthodentine<sup>2</sup>. Bases are rounded and cap-like with a row of five to six large foramina in the slightly concave foot. Cross-sections show osteodentine with a basal lamellar tissue, which directly abuts the dental membrane. The difference between the structure of the smaller posterior teeth (equivalent to type specimen BMNH (British Museum, Natural History) P.6540) and that of branchial denticles is still strong, contrary to one hypothesis on the origin of teeth<sup>17</sup>.

Woodward<sup>6</sup> diagnosed the taxon "*Diplodus*" problematicus on an isolated tooth (BMNH P.6540), concluding that the diplodont (xenacanth) tooth type was present by Early Devonian. Traquair<sup>8</sup>

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reassigned holotype and topotype specimens to *Doliodus*, with no doubt of its selachian nature, noting the succession of teeth. Since the early work and until recently<sup>2,7</sup>, however, the teeth have been erroneously assigned to the Acanthodii<sup>18</sup>. *D. problematicus* teeth are now reassigned to Chondrichthyes, with a structure similar to that of some xenacanthiforms, and included in the Omalodontida Turner 1997, known from shark teeth with a labially extended base<sup>2</sup>.

Until now, Pucapampella from the Late Emsian of South Africa<sup>4</sup> and Emsian?/Late Eifelian/Givetian of Bolivia<sup>10</sup> showed the oldest chondrichthyan cranium. The large braincase of Doliodus, in part and counterpart, is preserved from the precerebral fontanelle to just in front of the occipital region with the basal surface abutting the palatoquadrate (Fig. 2a, b). The precerebral fontanelle is prominent and large, similar to that of Tamiobatis<sup>9</sup> and Gladbachus<sup>13</sup>. We considered its presence as a putative elasmobranch synapomorphy<sup>10</sup> (neoselachian synapomorphy sensu Coates and Sequeira<sup>19</sup>); nonchondrichthyan taxa and holocephalans do not have a precerebral fontanelle<sup>10,19</sup>. The neurocranium of *Doliodus* is the oldest to possess this fontanelle. The Doliodus neurocranium has a moderately long otico-occipital region and shows similar proportions to Tamiobatis and Xenacanthus9. Postorbital processes are wider than in Pucapampella, although narrower than the total width of the otico-occipital unit, a purported plesiomorphic condition within chondrichthyans<sup>19</sup>. Poorly developed lateral otic processes are present, a character considered to be a chondrichthyan synapomorphy<sup>10</sup>. Evidence of a ventral otic fissure cannot be observed as preserved. Comparison of this feature to *Pucapampella*<sup>4,10</sup> is not possible. Posteriorly, an elongate median endolymphatic fossa is present; this character sets *Doliodus* as a chondrichthyan above *Pucapampella*<sup>10</sup>.

The mandibular, hyoid and branchial arches are preserved almost in life-position (Fig. 1), but the latter two are slightly displaced posteriorly. The right side is more difficult to interpret because of the overpositioning and lateral compression of the meckelian cartilage, basihyal, palatoquadrate, ceratohyal and anterior three ceratobranchials. The inverted U-shaped median basihyal is wide and constricted at the symphysis. The basihyal and basibranchials are separated by a gap. Two basibranchials are preserved; the posterior one being the largest. Four (perhaps five) pairs of elongated, slightly sigmoid ceratobranchials form the main part of the branchial apparatus. The visceral skeleton of *Doliodus* shows gross similarity to that of *Gladbachus*<sup>13</sup>, with the exception of the shape of the basibranchials and ceratobranchial IV.

Morphology of both pectoral fins is well preserved (Figs 1 and 3). Uniquely for chondrichthyans, fin-spines form the anterior margin of the pectoral fins in *D. problematicus*. Notably, the fin-spine of the articulated shark *Antarctilamna prisca* (CPC (Commonwealth





Figure 1 Partial articulated shark, *Doliodus problematicus* (NBMG 10127/1b,2,3). **a**, Specimen lying dorsal side up with the head at top and extending posteriorly to behind the pectoral fins. pfs, pectoral fin-spines. **b**, Map showing cartilage elements (black infill) and large areas of denticles (grey). mmd, location of mucous membrane denticles on counterpart; sym, symphysis; tth, area with *in situ* teeth; thf, *in situ* tooth family; mc, Meckel's cartilage; pq, palatoquadrate; bhy, basihyal; chy, ceratohyal;

bbra; anterior basibranchial; bbrp, posterior basibranchial; cbr, ceratobranchials (I–V?); sca, scapulocoracoid; pfs, pectoral fin-spines; rad, radials; sp, partial spines; denticle enlargements as preserved from branchial region (1), from pectoral fin (2) and from trunk region (3). Scale bar, 1 cm. Scales are separate for denticle enlargements.

Palaeontological Collection) 21187), which has been interpreted<sup>12</sup> as a dorsal spine, is suggestive of a pectoral fin-spine, being in the same position as the pectoral fin-spines described here. Until now, only dorsal fin-spines were known in basal chondrichthyans<sup>1,9</sup>. Paired, dermal pectoral fin-spines were previously known only in placoderms, acanthodians and the basal osteichthyan Psarolepis<sup>20</sup>. Their presence in Doliodus and perhaps Antarctilamna and the above-mentioned groups, suggests that it represents a gnathostome synapomorphy lost independently in Osteichthyes other than Psarolepis<sup>21</sup>, Placodermi and Chondrichthyes. The presence of large fin-spines associated with all fins except the caudal fin, or more specifically the presence of paired fin-spines, had been considered an acanthodian synapomorphy<sup>1</sup>. This character can no longer be considered an acanthodian synapomorphy and the tenuous monophyly of acanthodians is now supported by a single scale histology synapomorphy<sup>1</sup>.

The fin is aplesodic with radials extending half way to the margin (Figs 1 and 3). At least six (possibly seven) radials articulate on the lateral edge of a large basipterygial element (meso- or metapterygium); this feature is considered to be another chondrichthyan synapomorphy<sup>22</sup>. The area of dermal scales extends well lateral of the fin-spine distal tips, suggesting a large rounded pectoral fin. Left of the midline, near the posterior of the specimen, are small finspines that are furnished with hook-like denticles. These might be paired pelvic or intermediate spines, or parts of a collapsed dorsal fin-spine. Disarticulated fin-spines from the Campbellton Formation are identified as probable chondrichthyan and acanthodian<sup>6,8,23</sup>. Pectoral fin-spines on NBMG 10127 are closest in size and ornament to those that were originally named "Ctenacanthus" latispinosus<sup>23</sup> and subsequently reassigned to Climatius<sup>6</sup>, a climatiid acanthodian<sup>18</sup>. Isolated *Climatius latispinosus* spines, NBMG 9986 and 10017, are preserved with Doliodus teeth and scales, and spine NMC (Canadian Museum of Nature) 12002 includes a nearby patch







and patch of mucous membrane denticles with enlarged area outlined. **b** Map of neurocranium. pf, precerebral fontanelle; pop, postorbital process; ef, endolymphatic fossa; pq, palatoquadrate; mmd, mucous membrane denticles. Scale bar, 1 cm. **c**, Enlargement of mucous membrane denticles lining the inside of the mouth forward of the neurocranium. Scale bar, 0.5 cm.

of prismatic cartilage and branchial denticles. Further work is required to determine whether *D. problematicus* and *C. latispinosus* are synonymous.

*Doliodus* has rounded to polygonal, polyodontode mucous membrane denticles lining the inner upper palate and jaw edges (Fig. 2c) and rounded head denticles, multicuspid branchial denticles and ctenacanth-type complex trunk scales similar to those of *Antarctilamna* (Fig. 1), and thus is relatively advanced as compared with known purported Ordovician to Silurian sharks with simple placoid scales<sup>1,9</sup>. Behind the branchial region shagreen extends posteriorly, with scattered denticles and dorsal scales infilling the central body area. Trunk scale morphology is most like scales described from the (Pragian) Jauf Formation in Saudi Arabia<sup>24</sup>.

The Campbellton Formation<sup>25</sup>, with its rich flora<sup>26,27</sup> and terrestrial invertebrates<sup>28</sup>, yields vertebrates in the lower "Atholville beds"<sup>29</sup>. Miospores immediately below the articulated shark bed identify the *Emphanisporites annulatus–Camarozonotriletes sextantii* Assemblage Zone<sup>26</sup>, corresponding to early, to early late Emsian (*dehiscens* to *serotinus* Conodont Zones) age. *Doliodus* teeth<sup>6,8</sup> come from older beds at the base of the formation, perhaps near the 409-Myr-old Pragian/Emsian boundary<sup>5</sup>. The depositional environment has been considered fluvial<sup>25</sup>; however, fossil assemblages<sup>26–28</sup> suggest lagoonal and estuarine environments. Rare prasinophytes (tasmanids)<sup>26</sup> indicate a marine connection.

Discoveries of Early and Middle Devonian chondrichthyans from Gondwanan or neighbouring terranes have led to suggestions of a





Figure 3 Left pectoral fin of *Doliodus problematicus* (NBMG 10127/3). **a**, Left pectoral fin. **b**, Map of left pectoral fin. pfs, pectoral fin-spine; rad, radials I–VII(?); bas, basipterygial element (meso- or metapterygium). Scale bar, 1 cm.

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Gondwanan origin for sharks, although *D. problematicus* teeth presented a contradiction<sup>7</sup>. This specimen clearly places *D. problematicus* in Laurentia by the Early Devonian. Northern Gondwana and Laurentia were possibly close<sup>30</sup>, across a shallow shelf connecting north Gondwanan shoreline locales.

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- 1. Janvier, P. Early Vertebrates (Oxford Univ. Press, Oxford, 1996)
- Turner, S. in Recent Advances in the Origin and Early Radiation of Vertebrates (eds Arratia, G., Wilson, M. V. H. & Cloutier, R.) (Verlag Dr F. Pfeil, Muenchen, in the press).
- Mader, H. Schuppen und Z\u00e4hne von Acanthodiern und Elasmobranchiern aus dem Unterdevon Spaniens (Pisces). G\u00f6ttinger Arb. Geol. Pal\u00e4ont. 28, 1-58 (1986).
- Maisey, J. G. & Anderson, M. E. A primitive chondrichthyan braincase from the Early Devonian of South Africa. J. Vert. Paleontol. 21, 702–713 (2001).
- Tucker, R. D. et al. New U-Pb zircon ages and the duration and division of Devonian time. Earth Planet. Sci. Lett. 158, 175–186 (1998).
- Woodward, A. S. On the Lower Devonian fish-fauna of Campbellton, New Brunswick.. Geol. Mag. 9, 1–6 (1892).
- Long, J. A. & Young, G. C. Sharks from the Middle-Late Devonian Aztec Siltstone, southern Victoria Land, Antarctica. *Rec. West. Aust. Mus.* 17, 287–308 (1995).
- Traquair, R. H. Notes on the Devonian fishes of Campbelltown and Scaumenac Bay in Canada. *Geol. Mag.* 10, 145–149 (1893).
- Zangerl, R. in Handbook of Paleoichthyology Vol. 3A Chondrichthyes 1 (ed. Schultze, H.-P.) 1–114 (Gustav Fischer, Stuttgart, 1981).
- Maisey, J. G. in Major Events in Early Vertebrate Evolution. Palaeontology, Phylogeny, Genetics and Development. Systematics Assoc. Spec. Vol. Ser. 61 (ed. Ahlberg, P. E.) 263–288 (Taylor and Francis, New York, 2001).
- Janvier, P. & Suarez-Riglos, M. The Silurian and Devonian vertebrates of Bolivia. Bull. Inst. Fr. Etud. Andines 15, 73–114 (1996).
- Young, G. C. Devonian sharks from south-eastern Australia and Antarctica. *Palaeontology* 25, 817–843 (1982).
- Heidtke, U. H. J. & Krätschmer, K. Gladbachus adentatus nov. gen. et sp., ein primitiver Hai aus dem Oberen Givetium (Oberes Mitteldevon) der Bergisch Gladbach-Paffrath-Mulde (Rheinisches Schiefergebirge). Mainzer geowissenschaftliche Mitteilungen 30, 105–122 (2001).
- 14. Maisey, J. G. Heads and tails: a chordate phylogeny. Cladistics 2, 201-256 (1986).
- Reif, W.-E. in *Evolutionary Biology* Vol. 15 (eds Hecht, M. K., Wallace, B. & Prance, G. T.) 287–368 (Plenum, New York, 1982).
- 16. Compagno, L. J. V. Sharks of the Order Carcharhiniformes (Princeton Univ. Press, Princeton, 1988).
- Smith, M. M. & Coates, M. I. in *Major Events in Early Vertebrate Evolution. Palaeontology, Phylogeny, Genetics and Development.* Systematics Assoc. Spec. Vol. Ser. 61 (ed. Ahlberg, P. E.) 223–240 (Taylor and Francis, New York, 2001).
- Denison, R. in Handbook of Paleoichthyology Vol. 5 Acanthodii (ed. Schultze, H.-P.) 1–128 (Gustav Fischer, Stuttgart, 1979).
- Coates, M. I. & Sequeira, S. E. K. The braincase of a primitive shark. *Trans. R. Soc. Edinb. (Earth Sci.)* 89, 63–85 (1998).
- Zhu, M., Yu, X. & Janvier, P. A primitive fossil fish sheds light on the origin of bony fishes. *Nature* 397, 607–610 (1999).
- Zhu, M. & Schultze, H.-P. in *Major Events in Early Vertebrate Evolution. Palaeontology, Phylogeny,* Genetics and Development. Systematics Assoc. Spec. Vol. Ser. 61 (ed. Ahlberg, P. E.) 289–314 (Taylor and Francis, New York, 2001).
- Coates, M. I. & Sequeira, S. E. K. in *Major Events in Early Vertebrate Evolution. Palaeontology, Phylogeny, Genetics and Development.* Systematics Assoc. Spec. Vol. Ser. 61 (ed. Ahlberg, P. E.) 241–262 (Taylor and Francis, New York, 2001).
- Whiteaves, J. F. On some fossil fishes Crustacea & Mollusca from the Devonian rocks at Campbellton, NB, with descriptions of five new species. *Can. Nat.* 10, 93–101 (1881).
- Forey, P. L., Young, V. T. & McClure, H. A. Lower Devonian fishes from Saudi Arabia. Bull. Br. Mus. Nat. Hist. (Geol.) 48, 25–43 (1992).
- Williams, G. L., Fyffe, L. R., Wardle, R. J., Colman-Sadd, S. P. & Boehner, R. C. Lexicon of Canadian Stratigraphy Vol. VI Atlantic Region (Can. Soc. Petroleum Geologists, Calgary, 1985).
- Blieck, A. & Cloutier, R. in Palaeozoic Vertebrate Biochronology and Global Marine/Non-Marine Correlation (eds Blieck, A., Turner, S.) Cour. Forsch.-Inst. Senckenberg 223, 223–269 (2000)
- Correlation (eds Blieck, A., Iurner, S.) Cour. Forsch.-Inst. Senckenberg 223, 223–269 (200)
  Gensel, P. G. & Andrews, H. N. Plant Life in the Devonian (Praeger, New York, 1984).
- Gensel, F. G. & Andrews, H. N. Frank Life in the Devolution (Flagger, New York, 1984).
  Shear, W., Gensel, P. & Jeram, A. Fossils of large terrestrial arthropods from the Lower Devolian of
- Canada. Nature 384, 555–557 (1996).
- Dineley, D. L. & Williams, B. P. J. The Devonian continental rocks of the lower Restigouche River, Quebec. Can. J. Earth Sci. 5, 945–953 (1968).
- Keppie, J. D. & Ramos, V. A. in *Laurentia-Gondwana Connections Before Pangea* Geol. Soc. Am. Spec. Pap. 336 (eds Ramos, V. A. & Keppie, J. D.) 267–276 (Geol. Soc. Am., Colorado, 1999).

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# Temperature excludes N<sub>2</sub>-fixing heterocystous cyanobacteria in the tropical oceans

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Whereas the non-heterocystous cyanobacteria Trichodesmium spp. are the dominant N<sub>2</sub>-fixing organisms in the tropical oceans<sup>1</sup>, heterocystous species dominate N<sub>2</sub> fixation in freshwater lakes and brackish environments such as the Baltic Sea<sup>2</sup>. So far no satisfactory explanation for the absence of heterocystous cyanobacteria in the pelagic of the tropical oceans has been given, even though heterocysts would seem to represent an ideal strategy for protecting nitrogenase from being inactivated by O<sub>2</sub>, thereby enabling cyanobacteria to fix N<sub>2</sub> and to perform photosynthesis simultaneously. Trichodesmium is capable of N2 fixation, apparently without needing to differentiate heterocysts<sup>3</sup>. Here we show that differences in the temperature dependence of O<sub>2</sub> flux, respiration and N<sub>2</sub> fixation activity explain how Trichodesmium performs better than heterocystous species at higher temperatures. Our results also explain why Trichodesmium is not successful in temperate or cold seas. The absence of heterocystous cyanobacteria in the pelagic zone of temperate and cold seas, however, requires another explanation.

As primary production in vast areas of the oceans is predominantly controlled by the availability of nitrogen, biological N<sub>2</sub> fixation could overcome this limitation<sup>1</sup>. But N<sub>2</sub> fixation in the marine pelagic environment seems to be mainly restricted to (sub) tropical regions. The organisms responsible for most of the N2 fixation in the tropical oceans are Trichodesmium spp., filamentous non-heterocystous cyanobacteria that can form massive surface blooms<sup>4,5</sup>. Although free-living heterocystous cyanobacteria are reported to be present in the marine pelagic environment, their numbers are low and presumably they show very low growth rates<sup>6</sup>. This is notable because heterocystous cyanobacteria are considered to be better adapted to diazotrophic growth than are non-heterocystous species<sup>1,7</sup>, as heterocysts (differentiated cells enveloped by a glycolipid layer in which N<sub>2</sub> fixation takes place) are assumed to be effective in protecting the N2-fixing enzyme nitrogenase from inactivation by  $O_2$  (refs 8, 9).

The absence of heterocystous cyanobacteria in the marine pelagic environment contrasts strongly with their presence in freshwater lakes and brackish environments, where they can form dense blooms. Although heterocystous cyanobacteria can thrive in marine tropical systems, they are found mostly as epiphytes, in symbiosis with the planktonic diatom *Rhizosolenia* or in microbial mats<sup>10</sup>. But these specific environments are regularly oversaturated with  $O_2$ during the daytime, and therefore provide conditions that are different from those experienced by free-living organisms in pelagic systems. The expected higher  $O_2$  fluxes require a better protection of nitrogenase.

This leaves us with two main questions regarding the global distribution of  $N_2$  fixation. First, why are free-living heterocystous cyanobacteria not the dominant  $N_2$ -fixing organisms in the tropical oceans? Second, why are *Trichodesmium* spp. not able to thrive in marine, brackish or even freshwater environments in temperate and polar regions? Here, we propose that a glycolipid cell envelope, which acts as an effective diffusion barrier for  $O_2$  in heterocysts, does not provide an advantage in sea water at increased temperatures, and thus heterocystous cyanobacteria are out-competed by *Trichodesmium* spp. Our results also explain why *Trichodesmium*